# Effect of α-MSH and NPY on M1 Macrophages

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We used macrophages polarized to be pro-inflammatory M1 macrophages using lipopolysaccharide (LPS) and interferon- $\gamma$ . Macrophages produce cytokines which are small molecules that signal responses to other immune cells. IL-6 and TNF- $\alpha$  are pro-inflammatory cytokines while IL-10 is an anti-inflammatory cytokine.

In this study, we examined whether NPY or  $\alpha$ -MSH altered the production of specific cytokines including IL-6, TNF- $\alpha$ , and IL-10 to further understand if the suppression from these neuropeptides is by suppressing production of proinflammatory cytokines or by enhancing the production of anti-inflammatory cytokines to reduce inflammation.



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**Figure 1**: Macrophages are a type of white blood cell that are formed from monocytes and turn into macrophages when they enter tissue. The macrophages in this experiment were polarized through LPS and IFN- $\gamma$  to become M1 (proinflammatory). Figure 4. Immediately after adding the substrate Figure 5. 20 minutes after adding the substrate



**Figure 7**. Effect of  $\alpha$ -MSH and NPY on IL-6 levels



supported by the data from the TNF- $\alpha$  ELISA as the production of the pro-inflammatory cytokine TNF- $\alpha$  was significantly reduced in the NPY treated M1 macrophages compared to the untreated M1 macrophages.

- There was no anti-inflammatory cytokine, IL-10, made by the M1 macrophages or the M1 macrophages treated with the neuropeptides α-MSH and/or NPY.
- This suggests the mechanism by which the neuropeptides suppress the inflammation is by inhibiting the production of pro-inflammatory cytokines from M1 macrophages.

#### **Future Directions:**

- Analyze concentration levels of other proinflammatory cytokines such as IL-1β after the same neuropeptide treatments
- Increase the concentrations of neuropeptide treatments to see if there is greater inhibition or if other groups become significant

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#### **M1 Polarization:**

The macrophage cell line, RAW cells, were added into 10 wells within a 24-well plate. The groups were then treated with LPS and IFN- $\gamma$  to polarize into M1 macrophages.



**Figure 2:** This is a pictorial representation of the 24-well plate the cells were polarized and treated in. The groups were treated with their corresponding neuropeptides, A (cells), B (untreated), C (+ $\alpha$ -MSH+ NPY), D (+ $\alpha$ -MSH), and E (+NPY).

#### **Neuropeptide Treatment:**

Following 48 hours of incubation at 37°C and 5%  $CO_2$ , a Nitric Oxide (NO) assay was performed for culture supernatant for NO generation, a proinflammatory chemical to verify the polarization. Once confirmed, they were treated with their proper neuropeptides, either NPY and/or  $\alpha$ -MSH, or neither (negative and positive controls).

**ELISA:** Following a 24-hour incubation period at

The pro-inflammatory cytokine IL-6 was significantly suppressed (n=4, P  $\leq$  0.05) by the neuropeptide treatment of  $\alpha$ -MSH and NPY compared to the untreated (positive control) group.

#### **Figure 8**. Effect of $\alpha$ -MSH and NPY on TNF- $\alpha$ levels



### Figure 9. Table with significant results



## References

 Benque IJ, Xia P, Shannon R, Ng TF, Taylor AW. The Neuropeptides of Ocular Immune Privilege, α-MSH and NPY, Suppress Phagosome Maturation in Macrophages. Immunohorizons. 2018 Nov;2(10):314-323. doi: 10.4049/immunohorizons.1800049. PMID: 30613828; PMCID: PMC6319950.

 $37^{\circ}$ C and 5% CO<sub>2</sub>, the culture supernatant was collected and a sandwich ELISA was performed for either IL-6, IL-10, or TNF- $\alpha$  to measure the cytokine concentration. The concentration levels were then analyzed.



Figure 3. A sandwich
ELISA consists of a capture
antibody, the antigen, and a
detection antibody. It is
called a sandwich because
your antigen is bound
between the capture and
detection antibodies.



The pro-inflammatory cytokine TNF- $\alpha$  was significantly suppressed (n=4, P  $\leq$  0.05) by the neuropeptide treatment of NPY compared to the untreated (positive control) group. Zhou R, Caspi RR. Ocular immune privilege. F1000
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